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CONTROLLED ENVIRONMENT Food safety issues when growing greenhouse and CEA leafy greens

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I'll talk about ...



Food safety hazards in greenhouse/CEA systems



How research is informing food safety issues unique to CEA



Best Practices to bolster food safety



How does food safety in controlled environments differ from open field agriculture?



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Greenhouse and CEA Systems - advantages

- Optimal water, nutrient, light needs
- Control environmental conditions such as temp
- Reduced/eliminated weed pressure
- Reduced macrofauna problems/herbivory pressure
- Reduced plant disease
- Reduced risk of airborne hazards
- Reduced risks at harvesting





Photos: S. Micallef

Greenhouse and CEA Systems - food safety lens

Optimal water, nutrient, light needs

Control environmental conditions such as temp

Reduced/eliminated weed pressure?

• Reduced macrofauna problems/herbivory pressure

• Reduced plant disease

Reduced risk of airborne hazards

Reduced risks at harvesting

• Inputs, materials, equipment

Microbial water quality!

Possible temperature, air movement, RH control challenges

Altered macrofauna

Requires well thought out harvesting plan

Reduced/

microbial

diversity

altered

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Seeds

- Seeds can carry plant and human pathogens
- Storage considerations
- Seeds can attract wildlife
- Recalls and plans how to trace a crop affected by a recall
- Seed treatments



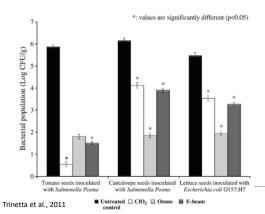


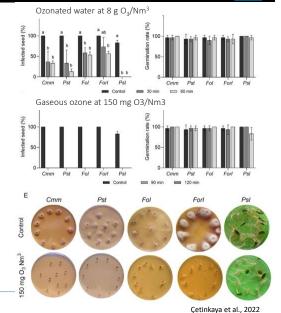
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Research: Seed



- · Disinfection can reduce pathogens
- · Hypochlorite, chlorine dioxide gas, gaseous ozone, ozonated water
- · Low concentration ozone can improve germination





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Substrates

- · Germination is achieved on solid substrate
 - Peat moss
 - Coco coir
 - Perlite
 - Vermiculite
 - Rockwool cubes
 - · Potting soil plugs







- Are they being reused?
- Can they be sanitized?
- Storage?



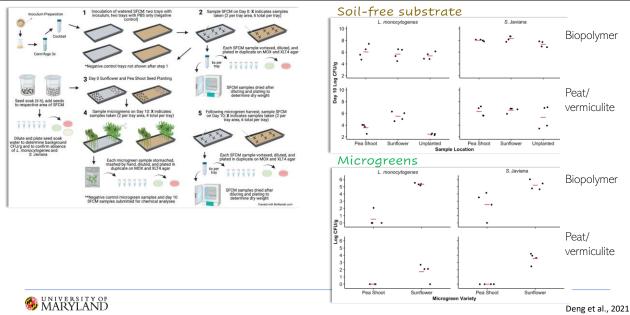




Photos: S. Micallef

Research: Enteropathogen survival in substrate





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Water: Why is water a risk?

- Provides ideal habitat (water, nutrients, temperature) for bacterial growth
- · Can serve as a reservoir
- Can serve as a vehicle, spreading bacteria through the system
- Splashed/leaked water can be tracked throughout facility
- Can lead to biofilm development



Water and Nutrient Solution

"Agricultural water ... intended to, or is likely to, contact the harvestable portion of covered produce"

- -sprinklers in high tunnel/greenhouse
- -irrigation drip lines
- -nutrient solution in NFT/deep water culture HP







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Photos: S. Micallef

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Open vs. closed hydroponic systems

- Open system:
 - Static/continuous contact with nutrient solution Roots may be in soil-free medium or in solution (e.g., deep water culture)
 - When crop cycle is complete, nutrient solution is drained as waste
- Closed system:
 - Nutrient solution is recirculating
 - (e.g., NFT)
 - Requires water treatment
 - Sand filters
 - UV illumination
 - · Antimicrobial agent such as Cl, PAA or ozone

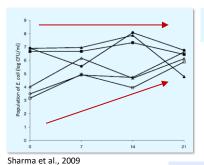




Photos: S. Micallef

Research: Pathogens in Nutrient Solution





E. coli O157:H7 in nutrient solution (pH 5.8) growing spinach at 25° C for 21 days

4 Log CFU inoculation of nutrient solution (pH 5.5-6.0) in NFT system growing butterhead lettuce at 12-30°C for 28 days

Salmonella Typhimurium		Listeria monocytogenes		
Time Post-Inoculation	Reservoir Nutrient Solution ¹	Channel Nutrient Solution	Reservoir Nutrient Solution ¹	Channel Nutrient Solution
1 h	2.21 a ²	2.19 ab	- 2	- 2
12 h	1.54 b	1.99 b	1.35 b ³	1.22 c
24 h	2.22 a	2.30 a	1.13 b	1.35 c
7 days	0.93 c	0.54 c	1.87 a	2.22 a
14 days	0.44 d	0.65 c	1.90 a	1.89 ab
21 days	0.38 d	0.53 c	1.34 b	1.60 bc
28 days	0.30 d	0.28 d	1.85 a	2.12 a
p value	<0.0001	<0.0001	<0.0001	<0.0001

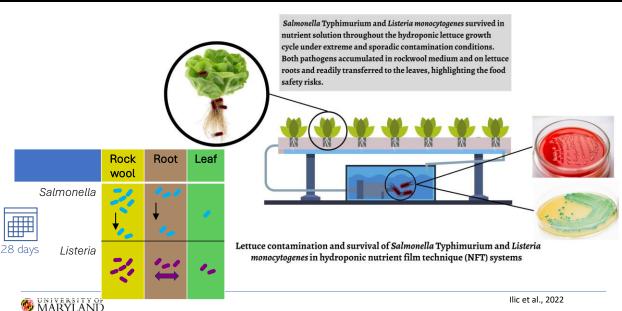
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Ilic et al., 2022

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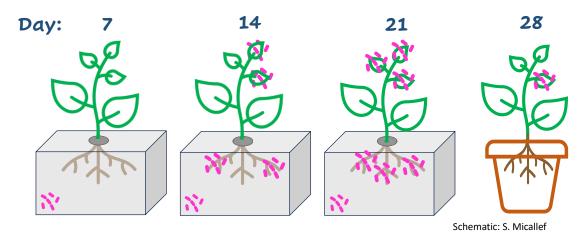
Research: Human Pathogens in NFT system





Research: Internalization into Crop





E. coli O157:H7 inoculated at low levels in nutrient solution. Surface sterilized spinach roots and shoots were tested after 7, 14 and 21 days. Plants were then transferred to soil and tested again.



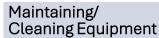
Sharma et al., 2009

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Food Contact Surfaces

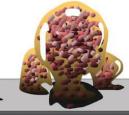
Biofilms

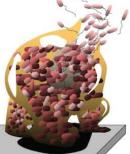
- 1 Reversible attachment of planktonic cells. (seconds)
- First colonizers become irreversibly attached. (second, minutes)
- Growth and cell division. (hours, days)
- Production
 of EPS and
 formation of
 water channels.
 (hours, days)
- 5 Attachment of secondary colonizers and dispersion of microbes to new sites. (days, months)



- Ponds
- Gutter systems
- Trays
- Rafts
- Harvest bins
- Work surfaces

Source: Open





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Wildlife and Pest control

- Rodents
- · Insects (aphids, thrips, whitefly), slugs, roaches
- · Occasional wildlife (birds, frogs)
- o Feces
- o Movement/cross-contamination
- o Herbivory
- > Monitor for signs
- Consider attractants, shelter
- Consider how trash, culled product is stored (inside and outside)
- ➤ Insect biological control ✓
- ➤ Traps ✓
- ➤ Poison X





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Harvest, Processing, Packing Workflow

- Direction of flow of people, crop, equipment
- Production water ≠ post-harvest water
- NFT vs deep water culture systems
- Harvest methods (roots or no roots?)











Photos: S. Micallef



In summary: Best Practices to consider

- In soil-less systems/hydroponics, consider seed disinfection
- Conduct Agricultural Water System Inspection (leaks, damage?)
- Do not allow for water to accumulate on floor or other areas
- Minimize contact between water and crop
- Consider water testing (generic E. coli)
- If recirculating water, consider best water treatment method
- Identify areas or steps where cross-contamination can occur
- Consider the workflow during production, during harvest (people, crop, tools)
- Develop a cleaning/sanitation plan
- Pest management plan
- Worker(s) training

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